

Role and Mechanism of AQP1 in the Transformation of Chronic Pain

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The mechanism of chronic pain transition is unclear. Some studies have revealed that aquaporin 1 (AQP1) is involved in chronic neuropathic pain, but its role in the mechanism of chronic pain transition has not been paid attention to. AQP1 is abundantly expressed in the small diameter, unmyelinated C fibers of dorsal root ganglion neurons, and co-located with nociceptant markers substance P and IB4. AQP1 is a new target for the treatment of pain. The signal transmission between macrophages and neurons promotes the sensitization of nociceptors. According to their polarization, they are divided into pro-inflammatory type (M1) and anti-inflammatory type (M2), and changing their ratio can regulate chronic neuropathic pain. Oxidative stress promotes chronic migraine. PI3K/Akt/HIF-1 α pathway has adaptive changes to oxidative stress and chronic hypoxia microenvironment, and intersects with p38MAPK pathway through downstream miRNAs/AQP1. This review summarizes the role of AQP1 in M1/M2 ratio and endothelial barrier function, and discusses that oxidative stress caused by early impairment of endothelial barrier function may be an important cause of delayed AQP1 expression enhancement. Oxidative stress, M1/M2 change and miRNAs/AQP1 signal in the microenvironment may respectively be important mechanisms and potential intervention targets of chronic pain transformation.

Keywords:chronic pain transformation; Oxidative stress; Endothelial barrier; miRNAs; AQP1; p38MAPK; PI3K; HIF-1 α

Aquaporins (AQPs) are a class of conserved transmembrane proteins expressed in various organ systems. They are cellular channels for bidirectional diffusion of water and small molecular compounds such as urea, lactate, hydrogen peroxide, and carbon dioxide. Human contains 13 kinds of aquaporins (AQP0-AQP12), which are widely distributed in various tissues and are involved in the pathological process of various non-infectious diseases, including renal insufficiency, nervous system diseases, skin diseases, metabolic syndrome and so on [1-3].

AQP1 is abundantly expressed in the small diameter, unmyelinated C-fiber neurons of Dorsal root ganglion (DRG).

AQP1 is abundantly distributed in the cell body and the periphery of neurons, and is densely expressed in the unmyelinated axons, myelinated axons and synaptic terminals. The co-localization of AQP1 with nociceptant markers substance P and IB4 is the anatomical basis for AQP1 to regulate some types of nociceptive signals [4-8]. DRG axon growth is impaired in AQP1-deficient mice, and the expression of AQP1 increases after sciatic nerve compression injury. AQP1 promotes axonal extension through water transport, which is a potential target for neuronal regeneration [6-7, 9]. AQP1 is a non-selective cation channel gated by cyclic guanosine monophosphate (cGMP) [10-12]. cGMP inhibitors reduce allodynia after chronic DRG compression injury in rats by reducing AQP1 levels [7,13]. The water permeability of neurons of AQP1 (-/-) mice and the perception of hot and cold pain induced by bradykinin, prostaglandin E2 and capsaicin are significantly reduced, which is a new target for pain treatment [7-8].

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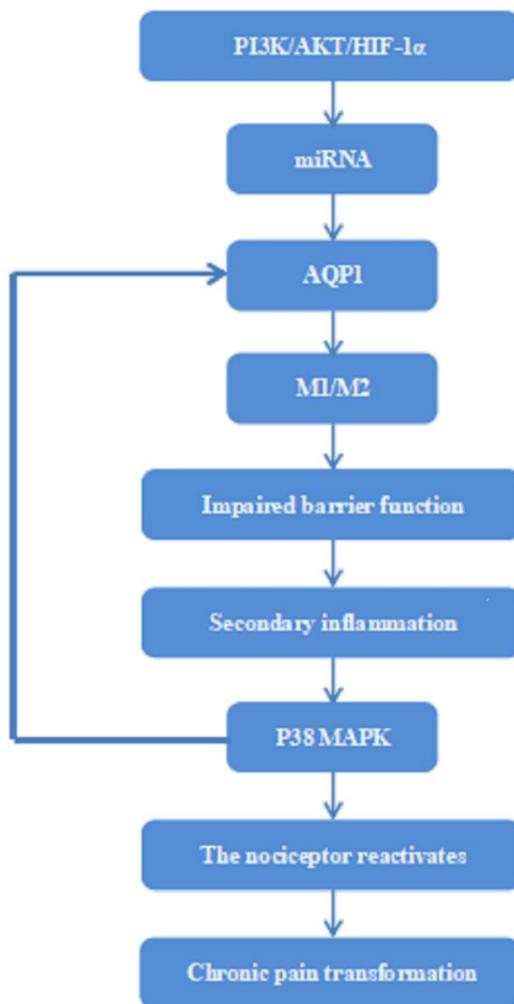
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MiRNAs regulate the transcription of most human protein-coding genes, and their dysfunction is related to the pathological process of many diseases [14], regulate the pathological changes of nervous system diseases [15], and have a huge impact on the development of neuropathic pain [16-17]. Ischemia and hypoxia reduce the level of AQP1, which is regulated by miRNAs subunits miR-214 and miR-133a-3p.1 [18-19]. *Mycobacterium leprae* is an intracellular pathogen that relies on the glutathione antioxidant system to control oxygen free radicals (ROS) and the REDOX state of

cells, thus changing the miRNA profile of leprosy patients, and promoting successful leprosy infection [20-22]. MiRNA participates in pathological processes such as skin dryness and loss of elasticity through AQP1 transcription loss in skin lesions, and is a new therapeutic target for leprosy pain [23], suggesting that oxidative stress microenvironment regulates AQP1 expression pattern, and miRNAs/AQP1 may be a potential mechanism for regulating chronic neuralgia signals.



P38 mitogen-activated protein kinase (p38 MAPK) pathway clears intracellular pathogens by affecting macrophage structure and function [24]. The transient inflammatory stimulation in the subacute phase after tissue incision is reactivated by p38 MAPK induced nociceptor, causing long-term nociceptive hypersensitivity and promoting the transformation of chronic pain, but the mechanism is unclear [25]. Pulmonary ischemia-reperfusion injury activates p38 MAPK to promote the increase of blood-air barrier permeability and macrophage infiltration by down-regulating the expression of AQP1, eNOS and VE-cadherin [26]. p38MAPK-AQP1 signal may be a new approach for targeted therapy of acute inflammatory injury [27]. These results suggest that secondary inflammation, oxidative stress and AQP1-mediated

macrophage and endothelial barrier dysfunction are important regulatory mechanisms and effectors of p38 MAPK, respectively. Combined with the literature report that "miR-495 overexpression activates p38MAPK to promote mouse osteoblast proliferation and differentiation by inhibiting AQP1 [28]", it is suggested that mirRNA-p38MAPK-AQP1 pathway may be a potential mechanism to regulate chronic inflammatory pain signaling.

Long-term and frequent migraine attacks increase oxidative stress events in the brain and promote the chronicity of migraine. Moreover, anti-migraine drugs have antioxidant effects [29-30], suggesting that oxidative stress microenvironment plays an important role in the transformation of chronic pain. Phosphatidylinositol 3-

kinase (PI3K)/protein kinase B (Akt)/hypoxia-inducible factor 1 α (HIF-1 α) pathway is involved in various biological processes such as oxidative stress and glycolysis [31-33], and activation of PI3K/Akt can reduce AQP1 levels [34]. HIF-1 α regulates key functions including cell migration, ROS production, pH [35] and is a major cause of intervertebral disc degenerative pain [36-37]. The level of AQP1 in DRG of rats with intervertebral disc degeneration pain increased significantly within 14 days [38], and the level of AQP1 at the site of spinal cord injury continued to increase for 11 months, and the delayed enhanced expression pattern was not related to local hypertonicity. Melatonin, an antioxidant, inhibits mechanical hyperalgesia in rats by reducing the level of AQP1 [39]. Low expression of AQP1 in the early stage of ischemia reperfusion promotes high microvascular permeability and high expression of HIF-1 α , and chronic hypoxia leads to high expression of AQP1 in the 7-14 days after injury [40]. These results suggest that oxidative stress microenvironment leads to delayed enhancement of AQP1 expression in subacute and chronic phase after injury, which induces nociceptor reactivation and long-term nociceptive hypersensitivity. PI3K/Akt/HIF-1 α may promote chronic inflammatory pain transformation through AQP1.

Loss of AQP1 leads to intercellular connectivity disorders, paracellular leakage, edema and macrophage infiltration in peripheral areas, neuroinflammation, and pain disorders [41-42]. Endothelial permeability was significantly increased in neuron-rich areas 7 days after nerve compression injury, but no further increase was observed 7 days later, but macrophage migration and nerve sensitivity were increased [43]. Endothelial barrier dysfunction leads to secondary damage such as inflammation induced by impaired peripheral blood supply [44], which further worsens the primary lesion [45]. These results suggest that early impairment of endothelial barrier function induces secondary inflammatory reactions such as oxidative stress and chronic hypoxia in damaged tissues in subacute and chronic stages after injury, which may be an important mechanism for the delayed enhancement of AQP1 expression, and may also be an important mechanism for the reactivation of nociceptors induced by activation of p38MAPK.

Signaling between macrophages and neurons promotes axonal sprouting and nociceptor sensitization [46-47]. According to their polarized state, macrophages can be divided into resting state (M0), proinflammatory type (M1) and anti-inflammatory type (M2). M1 produces proinflammatory cytokines, which promote sensitization of nociception, and M2 produces anti-inflammatory cytokines, which play multiple roles in the induction and resolution of inflammation. M2 releases a large number of opioid peptides including enkephalin and β -endorphin to control pain [48], and changing the M1/M2 ratio can alleviate chronic neuralgia in mice [49]. Xanthine oxidoreductase promotes oxidative stress and M1 polarization of sciatic nerve [50], and targeting the mitochondrial antioxidant peptide SS-31

inhibits M1 polarization and proinflammatory cytokine release by reducing ROS production in macrophages [51], suggesting that oxidative stress regulates M2/M1. Activation of PI3K/AKT promotes M1 polarization and induces inflammation [52-53]. HIF-1 α regulates immune metabolism and REDOX homeostasis of macrophages [54-55]. AQP1 relies on PI3K to promote M1 migration and M2/M1 change [56]. The activation of PI3K in AQP1siRNA mouse macrophages leads to the decrease of M2 [57], suggesting that there were positive and negative feedback effects between AQP1 and PI3K. PI3K/Akt/HIF-1 α /AQP1 regulates immune metabolism of macrophages and M2/M1 ratio. Combined with the literature reports that "HIF-1 α /miRNAs is a new regulatory mechanism for pathological processes such as myocardial fibrosis and asthma [58-60]", it is suggested that PI3K/Akt/HIF-1 α /miRNAs/AQP1 pathway may be a potential mechanism for regulating chronic inflammatory pain.

This article reviews the role of AQP1 in the change of M2/M1 ratio and endothelial barrier function, and discusses that the early damage of endothelial barrier leads to oxidative stress and dynamic changes of M2/M1 in the microenvironment, inducing delayed AQP1 expression enhancement, which may be an important mechanism for the transition of chronic pain. Since miRNAs/AQP1 is a crossing point between PI3K/Akt/HIF-1 α pathway and p38MAPK pathway, it is speculated that miRNAs/AQP1 may be a potential intervention target for the transformation of chronic pain.

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Reference

1. Jana D R Schmidt, Philipp Walloch, Bastian Höger, et al. Aquaporins with lactate/lactic acid permeability at physiological pH conditions. *Biochimie*. 2021 Feb 10; S0300-9084(21)00030-4.
2. Abul Kalam Azad, Topu Raihan, Jahed Ahmed, et al. Human Aquaporins: Functional Diversity and Potential Roles in Infectious and Non-infectious Diseases. *Front Genet*. 2021 Mar 16; 12:654865.
3. Georgios Tsiavaliaris, Fabian Ite, Kristina Hedfalk, et al. Low CO₂ permeability of cholesterol-containing liposomes detected by stopped-flow fluorescence spectroscopy. *FASEB J*. 2015 May; 29(5):1780-93.

4. Marios C Papadopoulos, Alan S Verkman. Aquaporin water channels in the nervous system. *Nat Rev Neurosci.* 2013 Apr;14(4):265-77.
5. Shannon D Shields, Javier Mazario, Kate Skinner, et al. Anatomical and functional analysis of aquaporin 1, a water channel in primary afferent neurons. *Pain.* 2007 Sep;131(1-2):8-20.
6. Hua Zhang, Alan S Verkman. Aquaporin-1 water permeability as a novel determinant of axonal regeneration in dorsal root ganglion neurons. *Exp Neurol.* 2015 Mar; 265:152-9.
7. Hua Zhang, Alan S Verkman. Aquaporin-1 tunes pain perception by interaction with Na(v)1.8 Na⁺ channels in dorsal root ganglion neurons. *J Biol Chem.* 2010 Feb 19;285(8):5896-906.
8. Kotaro Oshio, Hiroyuki Watanabe, Donghong Yan, et al. Impaired pain sensation in mice lacking Aquaporin-1 water channels. *Biochem Biophys Res Commun.* 2006 Mar 24;341(4):1022-89. Yasemin Kaya, Umut Ozsoy, Necdet Demir, et al. Temporal and spatial distribution of the aquaporin 1 in spinal cord and dorsal root ganglia after traumatic injuries of the sciatic nerve. *Childs Nerv Syst.* 2014 Oct;30(10):1679-86.
9. Jinxin V Pei, Sabrina Heng, Michael L De Ieso, et al. Development of a Photoswitchable Lithium-Sensitive Probe to Analyze Nonselective Cation Channel Activity in Migrating Cancer Cells. *Mol Pharmacol.* 2019 May;95(5):573-583.
10. Mohamad Kourghi, Michael L De Ieso, Saeed Nourmohammadi, et al. Identification of Loop D Domain Amino Acids in the Human Aquaporin-1 Channel Involved in Activation of the Ionic Conductance and Inhibition by AqB011. *Front Chem.* 2018 Apr 27; 6:142.
11. Jinxin V Pei, Mohamad Kourghi, Michael L De Ieso, et al. Differential Inhibition of Water and Ion Channel Activities of Mammalian Aquaporin-1 by Two Structurally Related Bacopaside Compounds Derived from the Medicinal Plant *Bacopa monnieri*. *Mol Pharmacol.* 2016 Oct;90(4):496-507.
12. Hui Wei, Wen-Shuang Gao, Lei Qi, et al. Effect of cGMP-activated aquaporin 1 on TRPV4 in rats with allodynia induced by chronic compression of the dorsal root ganglion. *Neurosci Lett.* 2020 Jan 18; 716:134630.
13. Anna L Malinowska, Artur Laski, Jonathan Hall. Design and Application of Mini-libraries of the miRNA Probes for an Efficient and Versatile miRNA-mRNA Cross-linking. *Chemistry.* 2021 May 17.
14. Masako Nakano, Mineko Fujimiya. Potential effects of mesenchymal stem cell derived extracellular vesicles and exosomal miRNAs in neurological disorders. *Neural Regen Res.* 2021 Dec;16(12):2359-2366.
15. Peng Chen, Chen Wang, Dongsheng Lin, et al. Identification of Slc6a19os and SOX11 as Two Novel Essential Genes in Neuropathic Pain Using Integrated Bioinformatic Analysis and Experimental Verification. *Front Neurosci.* 2021 Jan 28; 15:627945.
16. Yuanyuan Li, Chengyu Yin, Boyu Liu, et al. Transcriptome profiling of long noncoding RNAs and mRNAs in spinal cord of a rat model of paclitaxel-induced peripheral neuropathy identifies potential mechanisms mediating neuroinflammation and pain. *J Neuroinflammation.* 2021 Feb 18;18(1):48.
17. Arkady Rutkovskiy, Marte Bliksøen, Vigdis Hillestad, et al. Aquaporin-1 in cardiac endothelial cells is downregulated in ischemia, hypoxia and cardioplegia. *J Mol Cell Cardiol.* 2013 Mar; 56:22-33.
18. Jiang Y, Ma R, Zhao Y, et al. MEF2C/miR-133a-3p.1 circuit-stabilized AQP1 expression maintains endothelial water homeostasis. *FEBS Lett.* 2019 Sep;593(18):2566-2573.
19. Rychelle Clayde Affonso Medeiros, Karina do Carmo de Vasconcelos Girardi, Fernanda Karlla Luz Cardoso, et al. Subversion of Schwann Cell Glucose Metabolism by *Mycobacterium leprae*. *J Biol Chem.* 2016 Oct 7;291(41):21375-21387.
20. Amit Kumar Singh, Mrinmoy Ghosh, Vimal Kumar, et al. Interplay between miRNAs and *Mycobacterium tuberculosis*: diagnostic and therapeutic implications. *Drug Discov Today.* 2021 Jan 23; S1359-6446(21)00050-7.
21. Pablo Pinto, Moisés Batista da Silva, Fabiano Cordeiro Moreira, et al. Leprosy piRnome: exploring new possibilities for an old disease. *Sci Rep.* 2020 Jul 28;10(1):12648.
22. Claudio Guedes Salgado, Pablo Pinto, Raquel Carvalho Bouth, et al. miRNome Expression Analysis Reveals New Players on Leprosy Immune Physiopathology. *Front Immunol.* 2018 Mar 9;9:463.
23. Ligu Mi, Yan Wang, Hui Xu, et al. PRAK Promotes the Pathogen Clearance by Macrophage Through Regulating Autophagy and Inflammation Activation. *Front Immunol.* 2021 Apr 16; 12:618561.
24. Matsuda M, Oh-Hashi K, Yokota, Sawa T, et al. Acquired Exchange Protein Directly Activated by Cyclic Adenosine Monophosphate Activity Induced by p38 Mitogen-activated Protein Kinase in Primary Afferent Neurons Contributes to Sustaining Postincisional Nociception. *Anesthesiology.* 2017 Jan;126(1):150-162.
25. Wang T, Liu C, Pan LH, et al. Inhibition of p38 MAPK Mitigates Lung Ischemia Reperfusion Injury by Reducing Blood-Air Barrier Hyperpermeability. *Front Pharmacol.* 2020 Dec 11; 11:569251.
26. Bohui Li, Chunmei Liu, Kaihong Tang, et al. Aquaporin-1 attenuates macrophage-mediated inflammatory responses by inhibiting p38 mitogen-activated protein kinase activation in lipopolysaccharide-induced acute kidney injury. *Inflamm Res.* 2019 Dec;68(12):1035-1047.
27. Lei Zhu, Zun-Wen Lin, Gang Wang, et al. MicroRNA-495 downregulates AQP1 and facilitates proliferation and differentiation of osteoblasts in mice with tibial fracture through activation of p38 MAPK signaling pathway. *Sci Rep.* 2019 Nov 7;9(1):16171.
28. Kae M Pusic, Lisa Won, Richard P Kraig, et al. IFN γ -Stimulated Dendritic Cell Exosomes for Treatment of Migraine Modeled Using Spreading Depression. *Front Neurosci.* 2019 Sep 3; 13:942.
29. Patrizia Ferroni, Piero Barbanti, David Della-Morte, et al. Redox Mechanisms in Migraine: Novel Therapeutics and Dietary Interventions. *Antioxid Redox Signal.* 2018 Apr 20;28(12):1144-1183.
30. Xiangli Liu, Lidan Liu, Keyan Chen, et al. Huaier shows anti-cancer activities by inhibition of cell growth, migration and energy metabolism in lung cancer

- through PI3K/AKT/HIF-1 α pathway. *J Cell Mol Med.* 2021 Feb;25(4):2228-2237.
31. Lu Yiming, Han Yanfei, Yin Hang, et al. Cadmium induces apoptosis of pig lymph nodes by regulating the PI3K/AKT/HIF-1 α pathway. *Toxicology.* 2021 Mar 15; 451:152694.
 32. Zhang-Ping Yu, Han-Qiao Yu, Jun Li, et al. Troxerutin attenuates oxygen-glucose deprivation and reoxygenation-induced oxidative stress and inflammation by enhancing the PI3K/AKT/HIF-1 α signaling pathway in H9C2 cardiomyocytes. *Mol Med Rep.* 2020 Aug;22(2):1351-1361.
 33. Linlin He, Nan Zhang, Lei Wang, et al. Quercetin inhibits AQP1 translocation in high-glucose-cultured SRA01/04 cells through PI3K/Akt/mTOR Pathway. *Curr Mol Pharmacol.* 2020 Sep 8.
 34. Asmat Ullah, Sze Wei Leong, Jingjing Wang, et al. Cephalomannine inhibits hypoxia-induced cellular function via the suppression of APEX1/HIF-1 α interaction in lung cancer. *Cell Death Dis.* 2021 May 14;12(5):490.
 35. Bo Zhang, Qian Zhao, Yushi Li, et al. Moxibustion alleviates intervertebral disc degeneration via activation of the HIF-1 α /VEGF pathway in a rat model. *Am J Transl Res.* 2019 Sep 15;11(9):6221-6231.
 36. Shuai Chen, Xiang-Qian Fang, Qiang Wang, et al. PHD/HIF-1 upregulates CA12 to protect against degenerative disc disease: a human sample, in vitro and ex vivo study. *Lab Invest.* 2016 May;96(5):561-9.
 37. Dong Wang, Hao Pan, Hang Zhu, et al. Upregulation of nuclear factor- κ B and acid sensing ion channel 3 in dorsal root ganglion following application of nucleus pulposus onto the nerve root in rats. *Mol Med Rep.* 2017 Oct;16(4):4309-4314.
 38. O Nestic, J Lee, G C Unabia, et al. Aquaporin 1 - a novel player in spinal cord injury. *J Neurochem.* 2008 May;105(3):628-40.
 39. Liu M, Liu Q, Pei Y, et al. Aqp-1 Gene Knockout Attenuates Hypoxic Pulmonary Hypertension of Mice. *Arterioscler Thromb Vasc Biol.* 2019 Jan;39(1):48-62.
 40. Lisa Allnoch, Georg Beythien, Eva Leitzen, et al. Vascular Inflammation Is Associated with Loss of Aquaporin 1 Expression on Endothelial Cells and Increased Fluid Leakage in SARS-CoV-2 Infected Golden Syrian Hamsters. *Viruses.* 2021 Apr 8;13(4):639.
 41. Ying Hua, Xinxin Ying, Yiyu Qian, et al. Physiological and pathological impact of AQP1 knockout in mice. *Biosci Rep.* 2019 May 14;39(5): BSR20182303.
 42. Lux TJ, Hu X, Ben-Kraiem A, Blum R, Chen JT, Rittner HL. Regional Differences in Tight Junction Protein Expression in the Blood-DRG Barrier and Their Alterations after Nerve Traumatic Injury in Rats. *Int J Mol Sci.* 2019 Dec 31;21(1):270.
 43. M H Sheikh, S M Henson, R A Loiola, et al. Immunometabolic impact of the multiple sclerosis patients' sera on endothelial cells of the blood-brain barrier. *J Neuroinflammation.* 2020 May 9;17(1):153
 44. Reinhold AK, Rittner HL. Characteristics of the nerve barrier and the blood dorsal root ganglion barrier in health and disease. *Exp Neurol.* 2020 May; 327:113244.
 45. Christine Mary Barry, Dusan Matusica, Rainer Viktor Haberberger. Emerging Evidence of Macrophage Contribution to Hyperinnervation and Nociceptor Sensitization in Vulvodinia. *Front Mol Neurosci.* 2019 Aug 6; 12:186.
 46. Audrey Leung, Nicholas S Gregory, Lee-Ann H Allen, et al. Regular physical activity prevents chronic pain by altering resident muscle macrophage phenotype and increasing interleukin-10 in mice. *Pain.* 2016 Jan;157(1):70-79.
 47. Maria Pannell, Dominika Labuz, Melih Ö Celik, et al. Adoptive transfer of M2 macrophages reduces neuropathic pain via opioid peptides. *J Neuroinflammation.* 2016 Oct 7;13(1):262
 48. Muzamil Saleem, Brooke Deal, Emily Nehl, et al. Nanomedicine-driven neuropathic pain relief in a rat model is associated with macrophage polarity and mast cell activation. *Acta Neuropathol Commun.* 2019 Jul 5;7(1):108.
 49. Kazuhisa Takahashi, Hiroki Mizukami, Sho Osonoi, et al. Inhibitory effects of xanthine oxidase inhibitor, topiroxostat, on development of neuropathy in db/db mice. *Neurobiol Dis.* 2021 May 14;105392.
 50. Longcheng Shang, Haozhen Ren, Shuai Wang, et al. SS-31 Protects Liver from Ischemia-Reperfusion Injury via Modulating Macrophage Polarization. *Oxid Med Cell Longev.* 2021 Apr 13; 2021:6662156.
 51. Ye Li, Xiaomin Wang, Xiaoran Ma, et al. Natural Polysaccharides and Their Derivates: A Promising Natural Adjuvant for Tumor Immunotherapy. *Front Pharmacol.* 2021 Apr 14; 12:621813.
 52. Qi Wang, Song Wei, Haoming Zhou, et al. Hyperglycemia exacerbates acetaminophen-induced acute liver injury by promoting liver-resident macrophage proinflammatory response via AMPK/PI3K/AKT-mediated oxidative stress. *Cell Death Discov.* 2019 Jul 19; 5:119.
 53. Marco Tulio R Gomes, Erika S Guimarães, Fabio V Marinho, et al. STING regulates metabolic reprogramming in macrophages via HIF-1 α during Brucella infection. *PLoS Pathog.* 2021 May 14;17(5): e1009597.
 54. Ryan K Alexander, Yae-Huei Liou, Nelson H Knudsen, et al. Bmal1 integrates mitochondrial metabolism and macrophage activation. *Elife.* 2020 May 12;9: e54090.
 55. Donatienne Tyteca, Tomoya Nishino, Huguette Debaix, et al. Regulation of macrophage motility by the water channel aquaporin-1: crucial role of M0/M2 phenotype switch. *PLoS One.* 2015 Feb 26;10(2): e0117398.
 56. ChunMei Liu, BoHui Li, KaiHong Tang, et al. Aquaporin 1 alleviates acute kidney injury via PI3K-mediated macrophage M2 polarization. *Inflamm Res.* 2020 May;69(5):509-521.
 57. Shuang Liang, Ruihong Ning, Jingyi Zhang, et al. MiR-939-5p suppresses PM 2.5-induced endothelial injury via targeting HIF-1 α in HAECs. *Nanotoxicology.* 2021 May 3;1-15.
 58. Barry Garchow, Yvan Maque Acosta, Marianthi Kiriakidou. HIF-1 α and miR-210 differential and lineage-specific expression in systemic lupus erythematosus. *Mol Immunol.* 2021 May; 133:128-134.

59. Wang Feng,Zhao Ying,Fan Ke, et al.Apigenin suppresses TGF- β 1-induced cardiac fibroblast differentiation and collagen synthesis through the downregulation of HIF-1 α expression by miR-122-5p. Phytomedicine. 2021 Mar;83: 153481.

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